

doi.org/10.3114/fuse.2020.06.15

Tuber luomae, a new spiny-spored truffle species from the Pacific Northwest, USA

J. Eberhart^{1*}, J. Trappe², C. Piña Pérez³, G. Bonito⁴

¹Department of Crop and Soil Science, Oregon State University, Corvallis, Oregon, 97331, USA

²USDA, Forest Service, Forestry Sciences Laboratory, 3200 Jefferson Way, Corvallis, Oregon 97331, USA

³Department of Botany and Plant Pathology, Oregon State University, Corvallis, Oregon, 97331, USA

⁴Department of Plant Soil and Microbial Sciences, Michigan State University, East Lansing, Michigan 48823, USA

*Corresponding author: joyce.eberhart@oregonstate.edu

Key words:

new taxon
peridial structure
Pezizales
phylogeny
Rufum clade
truffle

Abstract: *Tuber luomae*, a new truffle species known only from the Pacific Northwest, USA, is distinguished by spiny, non-reticulate spores and a two-layered peridium — the outermost layer (pellis) consists of inflated, globose to subpolygonal cells and the inner (subpellis) of narrow hyphae. ITS sequence analyses show that it has phylogenetic affinity to other *Tuber* species in the Rufum clade. The only other members of the Rufum clade with a strongly developed peridiopellis of large, inflated cells are the southern European *T. malacodermum* and *T. pustulatum* and the northern Mexican *T. theleascum*. We find it interesting that this peridial structure that is uncommon in the Rufum clade has been found in geographically disjunct species.

Citation: Eberhart J, Trappe J, Piña Pérez C, Bonito G (2020). *Tuber luomae*, a new spiny-spored truffle species from the Pacific Northwest, USA. *Fungal Systematics and Evolution* 6: 299–304. doi: 10.3114/fuse.2020.06.15

Effectively published online: 25 June 2020

Corresponding editor: P.W. Crous

INTRODUCTION

In 1980 Dr. Dan Luoma collected six *Tuber* ascomata on Orcas Island of the San Juan Islands, located in the straits separating coastal northwestern Washington (USA) from southern Vancouver Island (Canada). Microscopic examination revealed that although this collection has spiny spores and macroscopic characteristics that place it within the Rufum clade (Bonito *et al.* 2010), it is distinguished by a pellis of subglobose to globose or subpolyhedral cells up to 35 µm broad. Three other *Tuber* collections with the same morphological features have been discovered. Two of the collections were in northwestern Oregon, consisting of one ascoma in Benton County in 1962 and two ascomata in Clackamas County in 1995; one collection in southwestern Oregon consisted of one ascoma in Jackson County in 2012. We here describe this new species, *Tuber luomae*, and use ITS rDNA sequence data to place it within the Rufum clade of the genus *Tuber*.

MATERIALS AND METHODS

Specimens

Spiny-spored *Tuber* species, collected by raking (Weber *et al.* 1997) during the past four decades, were examined in this study. When possible, data on color and morphology of fresh ascomata were recorded and the specimens photographed prior to air-drying. Sections of the type specimen of *Tuber luomae* were paraffin-embedded, sectioned by microtome, and stained with Safranin O/ Fast-green (Johansen 1940). The other specimens

were rehydrated in water and characterized anatomically by light microscopy. Spore measurements exclude the surface ornamentation of spines.

Phylogenetic analysis

Collections examined in this study and analyzed by molecular methods are presented in Table 1 along with their collection information and GenBank accession numbers (the Benton County collection had been preserved in an ethanol-formalin solution that had dried; reliable sequences were not obtainable).

Two procedures were used for molecular analysis. In the first, prior to DNA extraction, a small piece of previously unexposed gleba tissue was removed from the holotype (OSC 148707) collection with a sterile razor blade and pulverized with the aid of sterile sand, cubic zirconia beads and a Mini-Beadbeater (Biospec Products, Bartlesville, OK). DNA was extracted with 24:1 chloroform:isoamyl alcohol and precipitated with isopropanol. The internal transcribed spacer region (ITS1, 5.8S nrDNA, and ITS2) and part of the nuclear ribosomal large subunit (LSU) locus were amplified with the universal fungal primer sets ITS5 – ITS4 and LROR – LR5 (Vilgalys & Hester 1990, White *et al.* 1990, Bertini *et al.* 1999). The PCR cocktail and protocol followed that of Healy *et al.* (2009). PCR products were stained with 1× SYBR Safe (Invitrogen, Carlsbad, CA) and visualized on a 1.0 % agarose gel buffered with TAE buffer. Gel electrophoresis products were viewed on a GelDoc XR imager (Bio-Rad Laboratories, Inc., Hercules, CA). Qiagen Quick-Clean columns were used to clean PCR products prior to sequencing.

A second set of molecular methods was applied to the paratype specimens from Clackamas (OSC 148706) and Jackson (OSC

Table 1. Collections examined in this study and analyzed by molecular methods along with their collection information and GenBank accession numbers (the Benton County collection had been preserved in an ethanol-formalin solution that had dried; reliable sequences were not obtainable).

Species	Collection ID ¹	Origin	GenBank Accession No.
<i>Tuber candidum</i>	SOC 727	USA: Oregon	AY830856
<i>Tuber ferrugineum</i>	n/a	n/a	AF132506
<i>Tuber huidongense</i>	IFS Y. Wang 89924	China	DQ478632
<i>Tuber indicum</i>	Tind-my02	China	DQ329365
<i>Tuber liaotongense</i>	Tsp-hr02	China	DQ478645
<i>Tuber luomae</i>	*JT6003	USA: Washington	FJ809887
	RB12-139	USA: Oregon	MH142475
	JT17457	USA: Oregon	MH142474
<i>Tuber lyonii</i>	JT5665	USA: Texas	FJ809883
<i>Tuber melanosporum</i>	1015	Israel	AF167096
<i>Tuber melosporum</i>	AH31737	Spain	JX402095
<i>Tuber nitidum</i>	BM105	Spain	FJ809885
<i>Tuber pustulatum</i>	JT32319	Spain	FJ809889
	AQUI 9725	Spain	MK211278
<i>Tuber quercicola</i>	SOC 733	USA: Oregon	AY918957
<i>Tuber regimontanum</i>	*ITC909	Mexico	EU375838
<i>Tuber rufum</i>	BOLO1506-1	Italy	AY112894
<i>Tuber rufum</i> var. <i>lucidum</i>	AZ2097	Italy	FJ809888
<i>Tuber spinoreticulatum</i>	*Uecker188	USA: Maryland	FJ748913
<i>Tuber taiyuanense</i>	IFS Y. Wang 610	China	DQ478637
<i>Tuber theleascum</i>	AQUI 9729	Mexico	MK211283
<i>Tuber wenchuanense</i>	FL-2013d strain HMAS 60239	China	JX267044

¹ Asterisk (*) denotes type collection.

151373) counties; the Sigma Extract-N-Amp™ (Sigma-Aldrich, St. Louis, MO) protocol was followed by use of 15 µL of extract solution and 30 µL of dilution solution. PCR was performed with the universal primer set ITS1-F – ITS4 (White *et al.* 1990, Gardes & Bruns 1993) or the reverse primer ITS4Tuber, which was designed to improve specificity for *Tuber* (CTC GAC TCG TAG AAG ACA CT, Bonito unpublished). PCR products were stained with ethidium bromide after gel electrophoresis. PCR products were cleaned with ExoSAP-IT® (Affymetrix, Santa Clara, CA) prior to sequencing.

Sanger sequencing was performed in both directions using a BigDye Terminator v. 3.1 Cycle Sequencing Kit (Applied Biosystems, Foster City, CA) with ITS1-F, ITS5 or LROR (forward) and ITS4, ITS4Tuber or LR5 (reverse). DNA sequences were determined on an ABI PRISM 3700 DNA Analyzer (Applied Biosystems, Foster City, CA). Forward and reverse sequences were assembled and Sequencher v. 4.0 (Gene Codes, Ann Arbor, MI) was used to manually edit sequences and ambiguous regions at the ends were removed. Both ITS and LSU sequences were queried against the NCBI public database GenBank with the BLASTN algorithm for comparison with other sequences and to verify that the sequences belonged to *Tuber*.

DNA sequences were aligned manually for phylogenetic analysis with MacClade v. 4.0 (Maddison & Maddison 2002) and ambiguously aligned regions were excluded. Phylogenetic inference was conducted on the ITS alignment with RAxML

computed through the CIPRES web portal (www.phylo.org) with a GTR model nucleotide substitution, and 1 000 bootstrap iterations. Outgroup taxa belonging to the *Melanosporum* clade were used because they are known to comprise the phylogenetic sister group to the *Rufum* clade within the genus *Tuber* (Bonito *et al.* 2010, 2013). Sequences produced with these procedures were deposited in GenBank under accession numbers FJ809887, MH142474 and MH142475, as was an LSU sequence not used in the alignment (FJ809812). Sequence alignments were deposited at TreeBASE under the accession number 22992.

RESULTS

Taxonomy

The description of *Tuber luomae* is based on the holotype. The three paratypes were carefully examined and did not differ in any significant details.

Tuber luomae Trappe, Eberhart, Piña Páez & Bonito, *sp. nov.* MycoBank MB807402. Fig. 1.

Etymology: In honor of distinguished mycologist Dr. Dan Luoma, collector of the holotype, for his many contributions to mycology.

Typus: USA, Washington, San Juan Co., Orcas Island, Camp Orkila, elev. 20 m, 31 Oct. 1980, D. Luoma, Trappe 6003 (**holotype** OSC 148707; ITS GenBank FJ809887, LSU GenBank FJ809812).

Description: *Ascomata* when dried 20–25 × 25–30 mm, irregular, the surface even or with prominent wrinkles. *Peridium* light orange brown, minutely verrucose with a reddish-brown pellis over a pallid subpellis. *Gleba* with light brown fertile tissue

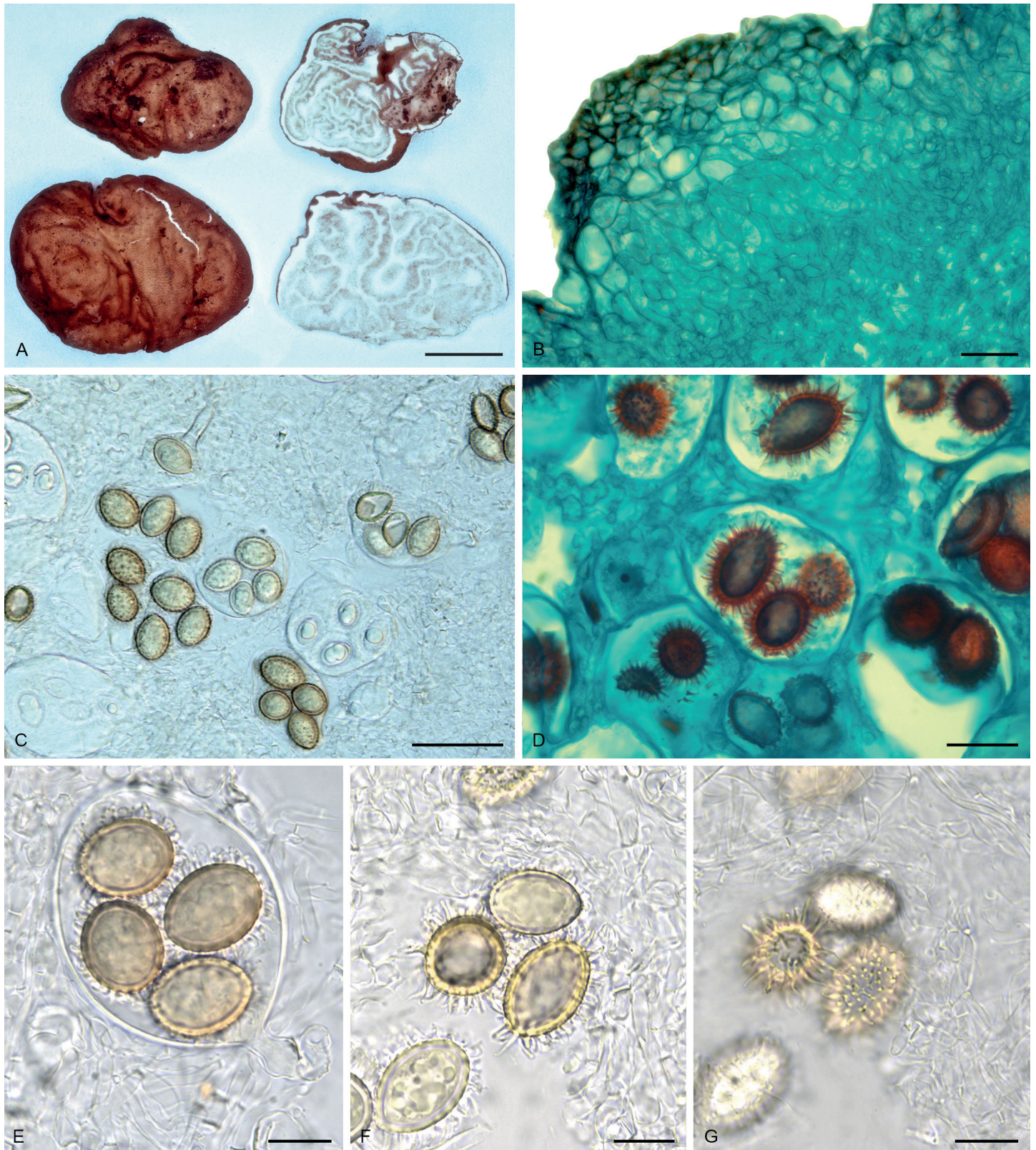


Fig. 1. Morphological characters of *Tuber luomae* (Holotype, OSC 148707). **A.** Ascocarp, surface and cross-section view. **B.** Outer peridium in cross section composed of inflated cells with inner peridium abruptly differentiated as interwoven hyphae, stained with Safranin O/ Fast-green. **C.** One- to four-spored asci. **D.** Spiny spores stained with Safranin O/ Fast-green. **E.** Four-spored ascus with distinctive basal hyphal stem. **F.** Spore ornamentation, highlighting the spines. **G.** Spores at two focal depths illustrating the spines and lack of connecting reticulum. Scale bars: A = 10 mm; B, D = 20 µm; C = 40 µm; E–G = 15 µm.

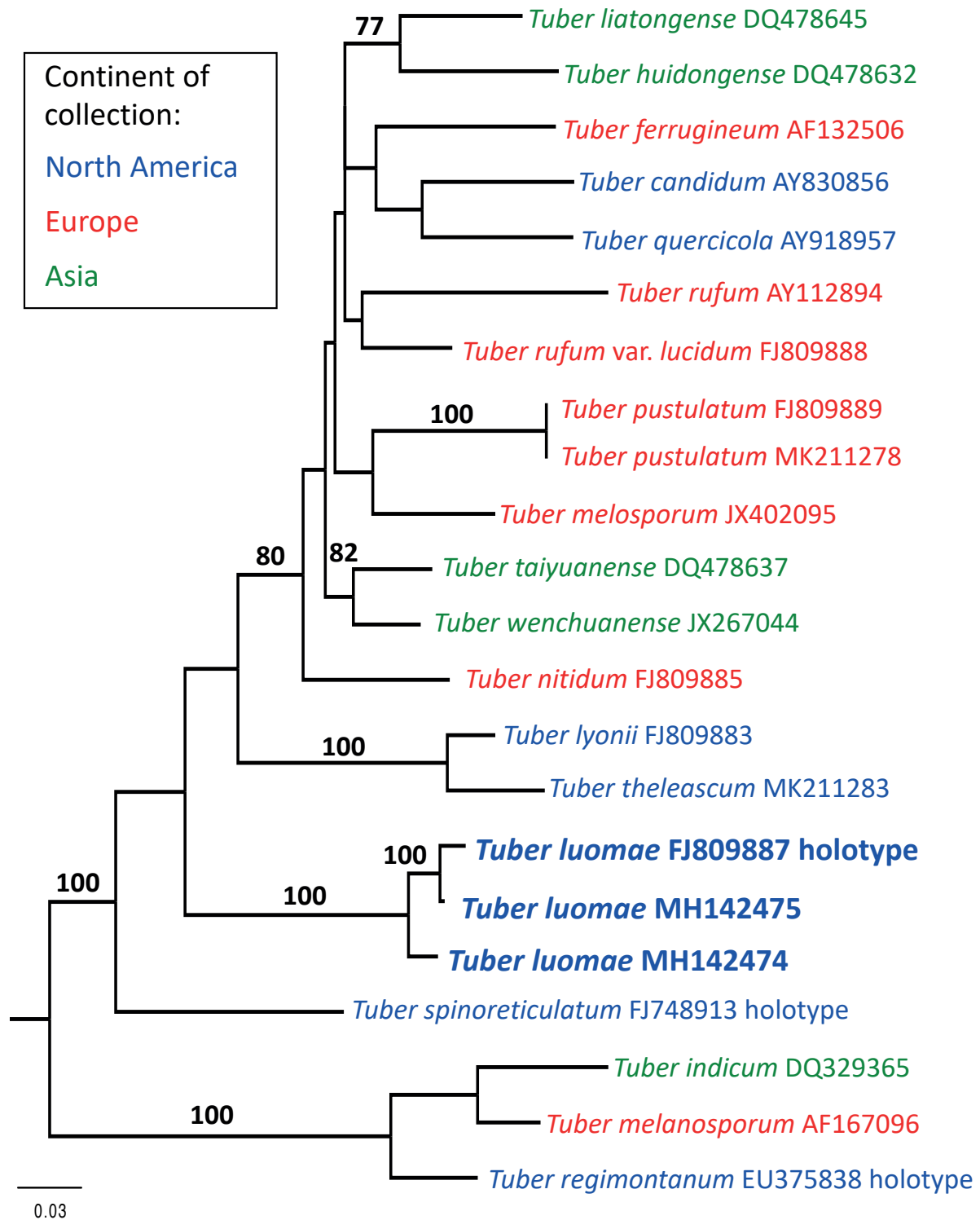


Fig. 2. Phylogenetic reconstruction of the *Tuber rufum* clade based on maximum likelihood analysis of the ITS rDNA region with 16 ingroup and 3 outgroup taxa (*T. melanosporum*, *T. indicum*, and *T. regimontanum*). The analysis was based on a GTR model of nucleotide substitutions and resulted in a most likely tree with a log likelihood score of $-\ln 4198.629$. Scale bar = 0.03 substitutions per site.

marbled with prominent white veins. *Odor* in the field mildly acrid. *Peridium* ± 500 μm thick, the outermost layer (pellis) ± 150 μm thick with crowded flattened warts 55–180 μm tall and 100–375 μm broad, of ellipsoid to subglobose or subpolyhedral cells 6–30(–38) μm broad, those of the outer ± 50 μm of the ascomata surfaces with yellow to yellowish brown walls 1–3 μm thick imparting an orange brown color to the tissue, below which the inflated cells are hyaline and walls mostly < 1 μm thick; subpellis ± 350 μm thick, gradually to abruptly differentiated from the

pellis as interwoven, hyaline, thin-walled hyphae 2–4(–8) μm broad. *Asci* hyaline, globose to ellipsoid or pyriform, 65–80 \times 40–60 μm excluding the basal hyphal stem, 10–38 \times 7–9 μm ; walls thin (< 1 μm) in youth, with age thickened to 2 μm . *Spores* 1–5/ascus, ellipsoid to broadly ellipsoid, ornamented with crowded, discrete hyaline spines 3–4 \times 1–1.5 μm . Spores in 1-spored asci (infrequent) 30–40 \times 24.5–35 μm ($Q = 1.18$ –1.45), 2-spored 25–33.5 \times 21–28 μm ($Q = 1.04$ –1.45), 3-spored 23–30 \times 18.5–23 μm ($Q = 1.21$ –1.32), 4-spored 20–26 \times 14–21 μm ($Q = 1.19$ –1.4),

5-spored (infrequent) $18\text{--}26 \times 15\text{--}21 \mu\text{m}$ ($Q = 1.2\text{--}1.31$); walls $\pm 2 \mu\text{m}$ thick, light yellow brown.

Distribution, habitat, and phenology: Cascade Mountains of far southwest Oregon and north on its western slopes as well as at the eastern slope of the Coast Ranges, thence to Washington's far north in the San Juan Islands; elevations range from 20 to 1 545 m. Habitats range from various mixtures and ages of *Pseudotsuga menziesii*, *Abies grandis*, and *Alnus rubra* to in one case, a partially harvested old-growth and in another, a pure stand of *Tsuga heterophylla*. The one collection in August was relatively immature, the other three were well matured.

Paratypes: **USA:** Oregon, Benton County, Rock Creek Park (abandoned) off State Hwy. 34, elev. 137 m, in soil, 24 Aug. 1962, *R. Benjamin* No. 38, OSC 158256; Clackamas County, Hwy. 224, SW of Timber Lake, elev. 550 m, in soil, 1 Nov. 1995, *A. Beyerle* B316, OSC 148706 (Trappe 17457); Jackson Co., Conde Creek, elev. 1 545 m, in soil, 4 Nov. 2012, *R. Brock* RB12-139, OSC 151373.

DISCUSSION

It is now evident from recently published molecular studies on *Tuber* and new species descriptions that *Tuber* is even more diverse than previously suspected (Bonito *et al.* 2010, Fan *et al.* 2011, 2012a, b, 2013, Alvarado *et al.* 2012a, Guevara *et al.* 2013, Zambonelli *et al.* 2016). In their assessment of the molecular diversity of *Tuber*, Bonito *et al.* (2010) and Healy *et al.* (2016) reported that the Rufum clade is among the more diverse in the genus and includes numerous cryptic undescribed species.

The Rufum clade is distributed across the Northern Hemisphere with centers of endemism in Europe, Asia, and North America (Bonito *et al.* 2013). While most species in this clade are characterized by spiny spores, some, such as *T. spinoreticulatum* and *T. lyonii*, have spiny spores with low reticulations (Uecker & Burdsall 1977, Trappe *et al.* 1996), whereas *T. liaotongense* has reticulate spores (Cao *et al.* 2011), and *T. melosporum* has smooth spores (Alvarado *et al.* 2012b).

Tuber luomae, known only from four collections in the Pacific Northwest (USA), is characterized by spiny-spores and a thick peridiopellis consisting of a pseudoparenchyma of inflated, subglobose to subpolygonal cells. Until recently, the only other described species in clade Rufum with a strongly pseudoparenchymatous pellis was *T. malacodermum* (Fischer 1923), known only from the holotype from Switzerland. Unlike *T. luomae*, it has a smooth peridiopellis, and the spines on its spores are occasionally to frequently connected by low ridges ('lines') on the spore surface, sometimes forming a partial reticulum. In addition, *T. malacodermum* has smaller asci but longer ascus stems than *T. luomae*. Despite their similar and distinctive peridial anatomy, they are readily distinguished by their spore ornamentation, ascus size, and ascus stem length.

Leonardi *et al.* (2019) examined the *T. malacodermum* holotype, as well as putative *T. malacodermum* ascomata collected in Spain, Corsica, and Mexico. Because morphological characters separated the type specimen from all the other collections, the authors concluded that the holotype is the only known collection of *T. malacodermum*. The type collection was too poorly preserved to yield DNA sequences, but molecular analysis of the newer species placed them into two previously

unidentified groups. The specimens from Spain and Corsica were named *T. pustulatum*, whereas the Mexican specimens were named *T. theleascum* (Leonardi *et al.* 2019). Molecular analyses by Leonardi *et al.* (2019) show that *T. luomae* is more closely related to the North American *T. theleascum* than to the European *T. pustulatum* (Fig. 2).

Tuber luomae resembles *T. pustulatum* in the structure of the peridium; both species have a two-layered peridium with an outermost layer composed of globose to angular cells and a subpellis of interwoven hyphae. They differ in the spore ornamentation; spines are shorter in *T. luomae* (up to $4 \mu\text{m}$ long) in comparison with *T. pustulatum* (up to $7 \mu\text{m}$ long). Moreover, the spines in *T. pustulatum* are united at the base, forming a reticulum, while the spines in *T. luomae* are discrete (Fig. 1F, G). *Tuber luomae* is also similar to *T. theleascum* in peridial structure, but differs in the thickness of the peridium (*T. theleascum* $160\text{--}250 \mu\text{m}$, *T. luomae* $\pm 500 \mu\text{m}$). They also differ in spore ornamentation, with spines often interconnected in *T. theleascum* but not connected in *T. luomae*.

These four species (*Tuber luomae*, *T. malacodermum*, *T. pustulatum* and *T. theleascum*), which have in common a peridiopellis of isodiametric cells rather than the narrow hyphae of all other known members of the clade, are derived from separate lineages. We find it interesting that this peridial structure that is uncommon in the Rufum clade has been found in geographically disjunct species (Fig. 1). *Tuber luomae* has phylogenetic affinity to other North American *Tuber* species in the Rufum clade. In the Pacific Northwest, other Rufum clade species (*T. candidum* and *T. quercicola*) are readily distinguished from *T. luomae* by having a peridiopellis of tightly interwoven hyphae that lack or have only widely scattered inflated cells. *Tuber luomae* can be regarded as rare, considering that hypogeous sporocarps (truffles) have been extensively sought by many dozens of collectors producing thousands of collections over more than a century of searching in western Washington and Oregon.

ACKNOWLEDGEMENTS

The authors thank Dan Luoma, Richard Benjamin, Adrian Beyerle, and Richard Brock for their collections of *T. luomae*. Microslides were sectioned and stained by Darr M. Duff. JT and GB were supported in this work through National Science Foundation awards 0641297 and 1946445. We thank Oregon State University for accessioning the collections of *Tuber luomae* discussed in this paper, Kyle Gervers for assistance in the OSC herbarium during times of limited access, and MycoBank for registering the new name.

Conflict of interest: The authors declare that there is no conflict of interest.

REFERENCES

- Alvarado P, Moreno G, Manjón LJ (2012a). Comparison between *Tuber gennadii* and *T. oligospermum* lineages reveals the existence of the new species *T. cistophilum* (Tuberaceae, Pezizales). *Mycologia* **104**: 894–910.
- Alvarado P, Moreno G, Manjón LJ (2012b). A new *Tuber* without spore ornamentation, *Tuber melosporum* comb. nov. *Boletín de la Sociedad Micología Madrid* **36**: 23–28.

- Bertini L, Amicucci A, Agostini D, *et al.* (1999). A new pair of primers designed for amplification of the ITS region in *Tuber* species. *FEMS Microbiology Letters* **173**: 239–245.
- Bonito G, Smith ME, Nowak M, *et al.* (2013). Historical biogeography and diversification of truffles in the *Tuberaceae* and their newly identified southern hemisphere sister lineage. *PLOS one* **8**: 1–15, e52765.
- Bonito G, Gryganskyi AP, Trappe JM, *et al.* (2010). A global meta-analysis of *Tuber* ITS rDNA sequences: species diversity, host associations and long-distance dispersal. *Molecular Ecology* **19**: 4994–5008.
- Cao JZ, Chi WD, Fan L, *et al.* (2011). Notes on the neotype of *Tuber taiyuanense*. *Mycotaxon* **116**: 7–11.
- Fan L, Cao JZ, Li Y (2013). A reassessment of excavated *Tuber* species from China based on morphology and ITS rDNA sequence data. *Mycotaxon* **124**: 155–163.
- Fan L, Cao JZ, Zheng ZH, *et al.* (2012a). *Tuber* in China: *T. microspermum* and *T. microspiculatum* spp. nov. *Mycotaxon* **119**: 391–395.
- Fan L, Hou CL, Cao JZ (2011). *Tuber sinoalbidum* and *T. polyspermum* - new species from China. *Mycotaxon* **118**: 403–410.
- Fan L, Hou CL, Li Y (2012b). *Tuber microverrucosum* and *T. huizeanum* - two new species from China with reticulate ascospores. *Mycotaxon* **122**: 161–169.
- Fischer E (1923). Zur Systematik der schweizerischen Trüffeln aus den Gruppen von *Tuber excavatum* und *rufum*. *Verhandlungen der Naturforschenden Gesellschaft in Basel* **35**: 34–50.
- Gardes M, Bruns TD (1993). ITS Primers with enhanced specificity for *Basidiomycetes* - Application to the identification of mycorrhizae and rusts. *Molecular Ecology* **2**: 113–118.
- Guevara G, Bonito G, Trappe JM, *et al.* (2013). New North American truffles (*Tuber* spp.) and their ectomycorrhizal associations. *Mycologia* **105**: 194–209.
- Healy RA, Bonito G, Guevara G (2009). The truffle genus *Pachyphloeus* in the U.S. and Mexico: phylogenetic analysis and a new species. *Mycotaxon* **107**: 61–71.
- Healy RA, Bonito GM, Smith ME (2016). A brief overview of systematics, taxonomy and ecology of the *Tuber rufum* clade. In: *True truffle (Tuber spp.) in the world* (Zambonelli A, Iotti M, Murat C, eds). Cham, Switzerland: Springer International Publishing. *Soil Biology* **47**: 125–136.
- Johansen, DA (1940). *Plantmicrotechnique*. McGraw-Hill Book Co., Inc., NewYork.
- Leonardi M, Paz-Conde A, Guevara G, *et al.* (2019). Two new species of *Tuber* previously reported as *Tuber malacodermum*. *Mycologia* **111**: 376–389.
- Maddison D, Maddison W (2002). *MacClade: Analysis of Phylogeny and Character Evolution*. Version 4.0. Sunderland: Sinauer Associates.
- Trappe JM, Jumpponen A, Cázares E (1996). NATS truffle and truffle-like fungi 5: *Tuber lyonii* (= *T. texense*), with a key to the spiny-spored species groups. *Mycotaxon* **60**: 365–372.
- Uecker FA, Burdsall HH (1977). *Tuber spinoreticulatum*, a new truffle from Maryland. *Mycologia* **69**: 626–630.
- Vilgalys R, Hester M (1990). Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several species of *Cryptococcus*. *Journal of Bacteriology* **172**: 4238–4246.
- Weber NS, Trappe JM, Denison WC (1997). Studies on western American *Pezizales*. Collecting and describing ascomata: macroscopic features. *Mycotaxon* **61**: 153–176.
- White, TJ, Bruns T, Lee S, Taylor J (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: *PCR Protocols: A guide to Methods and Applications* (Innis MA, Gelfand DH, Shinsky JJ, *et al.*, eds.), Academic Press, New York, USA: 315–322.
- Zambonelli A, Iotti M, Murat C, eds. (2016). *True truffle (Tuber spp.) in the world*. Cham, Switzerland: Springer International Publishing. *Soil Biology* **47**.